ENTIFIC RESEARC



DOI:10.32606/IJSR.V13.I1.00002

Received: 16-05-2022

Accepted: 17-08-2022

Publication: 31-08-2022 Original Research Article

Indian J.Sci.Res. 13 (1): 09-19, 2022

CHANGES OF PHYSIOCHEMICAL PROPERTIES OF SUNFLOWER OIL DUE TO ADDITION OF ANTIOXIDANT

ABDELHADI A. MOHAMMED^{a1}, MUTAZ M. A. ELSHEKHI^b AND AYMAN A. JACKNOON^c

^{ab}Department of Chemistry, Ahfad University for Women, Sudan

^cDepartment of Chemistry, Faculty of Science and Technology, Alneelain University, Sudan

ABSTRACT

This study was carried out in order to probe the extent of oxidative alterations in Sunflower Oil (SFO), subjected to light and dark storage conditions with, and without added vitamins, over a period of 16 weeks storage. The magnitude of oxidative changes was monitored by the periodical measurement of acid value, peroxide value, iodine value, Density and color throughout storage time. The acid value of sunflower oil increased from initial value 3.29 mg KOH/g to (3.56 and 4.80) mg KOH/g, the acid value for oil samples kept in light and dark with added vitamin C, vitamin E and equal amount mixed from vitamin C and E was decreased for 16 weeks. Like the acid value, the peroxide value of sunflower oil increased from 7.28 meg O₂/kg to (7.28 and 9.69) meq O₂/kg, for oil samples kept in the same conditions with that mentioned in acid value. Unlike the acid value, the Iodine value of sunflower oil decreased from 134.5 gI₂/100g oil to (98.53and 97.1) gI₂/100g, the initial density 0.9005 g/cm⁻¹ was increase with added antioxidants vitamins The density values was calculated after storage time and observing in the ranges recommended by standard for edible vegetable oils this is a results of density are the acceptability of oil quality. Like the density, the color of oils was not change because the oil is rich by the color pigments. However, the vitamins added of fresh sunflower oils increase the oxidation stability. Changes in acid value, peroxide value and Iodine value obtained shows that the oxidative deterioration levels of oils were different between storage conditions. The results of the present study show that light acts as a major catalyst in accelerating the development of rancidity in oils. Also, the addition of vitamins to oil can increase the oxidation stability of oils during storage. In conclusion, this study has been able to show storage in the light can affect oil stability and minimizes the potency of vitamins in oils, fats or fat-containing products. Therefore, storing in dark (packaging with material protect light) and supporting with antioxidants is the best way to maintain the quality of oils during storage and domestic uses.

KEYWORDS: Al-Jazeera Sunflower Oil, Vitamin C, Vitamin E, Antioxidants, Oxidation

Sunflower (Helianthus annuus L.) is one of the important oilseed crops grown throughout the world as a source of premium oil and dietary fiber that significantly contributes to human health (Khan et al., 2015). Sunflower seed contain a high amount of oil (40% to 50%) which is an important source of polyunsaturated fatty acid of potential health benefits (Monotti, 2004). Oil is a very important resource, much in demand everywhere in the world and is used in a variety of ways (Gizachew, 2020).

The physicochemical properties of fats and oils are effect by the degree of unsaturation, the length of the carbon chain and the type, quantity, distribution on the triglycerol and isomeric form of FA. The oils in sunflower seeds are rich of unsaturated fatty acids, rendering the seeds susceptible to oxidative rancidity this rancidity occurs after prolonged storage and is accelerated when the seeds are stored under inappropriate conditions. The oxidative rancidity affects the quality of both sunflower seeds and sunflower seed oil. (Meng, et al., 2019). The use of antioxidants as inhibitors of free radical autoxidation is of major importance in preserving

polyunsaturated lipids from oxidative deterioration (Frankel, 2005). An antioxidant can be classified as any substance that significantly delays or inhibits oxidation of a substrate.

Vitamin C or ascorbic acid is vitamin which function as a cofactor in many reaction due to its function as a reducing agent, this property of vitamin C makes it an important antioxidant, vitamin C is especially important as it also function to regenerate other antioxidants including alpha-tocopherol or vitamin E. (Sarkar et al., 2016). Vitamin E is found naturally in some foods, added to others, and available as a dietary supplement. "Vitamin E" is the collective name for a group of fat-soluble compounds with distinctive antioxidant activities .Naturally occurring vitamin E exists in eight chemical forms (alpha-, beta-, gamma-, and delta-tocopherol and alpha-, beta-, gamma-, and delta-tocotrienol) that have varying levels of biological activity. (Traber et al., 2006). Alpha- (or a-) tocopherol is the only form that is recognized to meet human requirements. (Traber, 2007).





Figure 1: Possible pathways of antioxidant mechanisms are illustrated

OBJECTIVES

Sunflower oil is considered as a concentrated source of energy for human beings and carriers of oilsoluble Vitamins which supply the essential fatty acids that are required for a wide range of biological and physiological functions. The oxidative changes during storage and domestic use make oils unsuitable for consumption. Oxidative stability is an important indicator of oil quality and shelf-life. This study is expected to increase awareness among the population regarding the influence of storage conditions and added vitamin E and C on the oxidation stability of domestically produced sunflower oil by adapting different oxidation detection methods. Furthermore, the study is important to see the quality and shelf life of domestically produced sunflower oils.

MATERIALS AND METHODS

Materials

All chemicals and reagents used in physicochemical analysis were analytical grade and were used with no further purification n-hexane, Ethanol, phenolphthalein, potassium hydroxide (KOH), glacial acetic acid, potassium iodide (KI), sodium thiosulfate (Na2S2O3), potassium bromide, starch were all obtained from Merck and Sigma Co. Chloroform was obtained from WINLAB LIMITED, United Kingdom. Vitamin E and C, was purchased from the SHUANGFENG Industrial China. Distilled water was used throughout the work.

Plants Collection

Fresh Sunflower seed (FSFS), obtained from Aljazeera state area in Sudan.

Equipment

The equipments used in this work are electronic balance (OHAUS, Switzerland), soxthlet apparatus, pycnometer, micro pipettes, burettes, conical flasks, thermometer, stirrer, stands and heating device.

Instruments

GC-MS spectrophotometry technique model (GC/MS –QP2010-Ultra) from japans (Shimadzu Company) with serial number 020525101565SA, The GC operating conditions were: column Rtx....length (30m)....Diameter (0.25mn), Detector mass spectrometer and carrier gas Helium with flow rate 1.61 ml/min,

Methods

General Methodology

The sample of sunflower seed was obtained from agricultural area in Sudan. Al-Jazeera state. Oil from the seed was obtained separately using solvent extraction method. Pure oil was separated using rotary evaporation. An oil sample from extract was investigated chemically and physically, the physical tests were density and color. And the chemical tests were acid value, peroxide value, iodine value. The oil sample was also subjected to GC-MS. Oil from sample was divided into four portions labelled A, B, C and D. portion A was the control sample, portion B was mixed with vitamin C, portion C was mixed with vitamin E and portion D was mixed with an equal amount of vitamin C and vitamin E. the same physical and chemical tests above were replicate to all portion at an interval of three times, average results were obtained. The same procedure was repeated to an oil samples stored in normal day light and temperature for eight weeks and sixteen weeks separately. Results were recorded and conclusion were obtained.

Extraction of Sunflower Oil

The sample of sunflower seed was extracted by soxthlet method. A total of 50 g sunflower seed sample was weighed and extracted with n-hexane in a Soxthlet Apparatus at a condensation rate of 5 or 6 points per second for 4 hours with 300 ml of hexane at a temperature of 70°C. The solvent was evaporated to dryness using a rotary evaporator at 40°C. (AOAC, 2005).

Physiochemical Analysis of Sunflower Oil

Determination Specific Gravity

Determined by the standard method of (AOAC, 2000).

Determination of Color

Color was determined according to standard method of (AOAC, 2000).

Determination Acid Value

The AV is the number of mg of KOH necessary to neutralize the free acid in 1 g of sample. Acid value was determined according to (Okpuzor, *et al.;* 2009).

The acidity is frequently expressed as free fatty acid for which calculation shall be.

Free fatty acids as oleic acid = 28.2 VN

Per cent by weight = W

Acid value = Percent fatty acid (as oleic) x 1.99

Determination Peroxide Value

Peroxide value (PV) was evaluated according to (AOCS, 2003).

Determination Iodine Value

Iodine value was determined according to the Hanus method as described in (A.O.A.C, 2000).

Preparation of Methyl Ester of Fatty Acid

The fatty acid methyl esters were prepared as described in the International Olive Council (IOOC, 2009).

Fatty Acid Profile

Fatty acid profile was analyzed by GC according to the method described by (IOOC, 2009).

Determination of Fatty Acid Methyl Easters

Gas chromatography has been used for the qualitative and quantitative analysis of the fatty acids reported in the relative area percentage, the GC/MS

technique model (GC/MS -QP2010-Ultra) from japans number (Shimadzu Company) with serial 020525101565SA and capillary column (Rtx -5ms -30m x 0.25mm x 0.25µm). The sample was injected by using split mode, helium as the carrier gas passed with flow rate 1.61 ml/min, the temperature program was started from 60°C with rate 10°C /min to 300°C as final temperature degree with 6 minutes hold time, the injection port temperature was 300°C the ion source temperature was 200 °C and the interface temperature was 250 °C. the sample was analyzed by using scan mode in the range of 40 to 500 m/z charge to ratio and the total run time was 30 min. Identification of components for the samples was achieved by comparing their retention times and mass fragmentation patents with those available in the library, the National Institute of Standards and Technology (NIST). The results were recorded.

Statistical Analysis

The statistical analysis was performed with the SPSS package software, version 20 (SPSS). Results were presented as means \pm standard deviation of the two triplicates of each experiment. Analysis of variance (ANOVA) was performed. Significant differences among the means (p < 0.05) were determined by Duncan's multiple tests.

RESULTS AND DISCUSSION

Characterization Analysis of Sunflower Oils

The chemical and Physical parameters are usually used for the identification of oils. Normally more than one character is determined so that the identification can be made with more assurance since the oils vary in their properties. The composition is not constant it depends upon certain factors such as climatic conditions, nature of soil, type of plant and variety of edible oil.

Physiochemical Characterizations Analysis

Physicochemical properties of oils are determined to know the quality, purity and identification. Characteristic properties are properties that depend on the nature of the oil. These are used to characterize oil, irrespective of location or sources of origin

The initial determined of Physiochemical characterization for sunflower oil samples have been made before the storage and antioxidants vitamin added Table 1. Shown the results of some characteristics of sunflower oil such as density, color, AV, FFA, PV and IV.

Sample	Density at 20°C (g/cm ⁻¹)	Color	Acid value (mg KOH/g oil)	Free Fatty Acid (%)	Peroxide value (meq O2/kg oil)	Iodine value (gI2/100g oil)
SFO	$0.9005 \pm (0.001)$	$12.2 \pm (0.07)$	$3.29 \pm (0.02)$	$1.65 \pm (0.01)$	$7.28 \pm (0.04)$	$134.50 \pm (0.71)$

Table 1: Physiochemical Characterization of Sunflower Oil

The above results show that density of sunflower oil are always greater than the refined oil. Therefore oil may provide more protection for human health than refined oil.

Changes in Specific Gravity

Specific gravity is considered as a good index of purity of oils, the increase in chain length of fatty acid present in oil tends to increase the specific gravity of oils. (Table 1) Shown the specific gravity value of oils samples were within the FAO/WHO standard for edible vegetable oils. The specific gravity of Al-Jazeera sunflower oil shows that the oil is less dense than water because the impurities are not present in oil.

Changes in Color

Change in color indicates the deterioration of oil due to oxidation. Color of oils depends upon the nature of coloring material like chlorophyll and carotene present in oil. Sunflower oil samples have pale yellow color indicating the presence of color pigments, so the color of oils was not change because the oil is rich by the color pigments, according the color of Al-Jazeera sunflower oils became stable.

Changes in Acid Value (AV)

AV is a measure of the FFAs in oil. Normally, FAs are found in the TAG form, however; during processing, the FAs may be hydrolyzed into FFA. Production of FFA is the best predictor of fat deterioration and the presence of FFA could be used to monitor the extent of oils abused (Atta et al., 2008). Table 1. Shows the lower AV found, the lower the level of FFA which results in increased oil quality. The initial AV of SFO presented in this study show a lower value than Codex Standard for Named Vegetable Oils (CODEX-STAN210-1999) (4.0 mg KOH/g). The observation of low initial AV for Al-Jazeera sunflower oil $(3.29 \pm 0.02 \text{ mg KOH/g of oil})$ indicates a low formation of FFAs. Probably, this is due to oil seed without moisture content. The acceptability level of virgin sunflower oils is below 4.0 mg KOH/g (measured in potassium hydroxide per gram). (Alimentarius, 1999).

Changes in Fatty Acid Composition of Oil

Change in free fatty acid values were determined for sunflower oils (Table.1). Although the initial free fatty acid value for the sample of the SFO was almost insignificantly different so the value was found in the sample.

Changes in Peroxide Value (PV)

Determination of peroxide value can give an idea about the early stages of oil oxidation. The PV indicates the level of oxidation during production and storage. One of the most important parameters that influence lipid oxidation is the degree of unsaturation of its FAs. When double bonds of unsaturated fats are oxidized, peroxides are among the oxidation products formed. The peroxide values for the fresh oil was very low which indicate the high quality of the SFO used in this work. Change in PV without storage and antioxidant vitamins were shown in Table. 1.

Peroxides are responsible for the taste and odor of rancid fats, their concentration as represented by the PV is often useful in assessing the extent to which the rancidity has advanced. Rancid taste often begins to be noticeable when the PV is above 20 meq O2/kg (Food Adulteration, 1954). At the beginning of the experiment, the PV of SFO was (7.28 ± 0.035 meq O2/kg. Which is less than 10 meqO2/Kg, and therefore within the acceptable value range for fresh oil. This value within the range considered as satisfactory and in agreement with the maximum Codex standard PV (15 meq O2/Kg). For virgin vegetable oil. (Alimentarius, 1999).

Changes Iodine Value (IV)

The iodine value is a measure of the unsaturation of the oils, it is one of the parameters used to measure the oil quality, Table 1. Demonstrates the initial iodine value of SFO is (134.50 gI2/100g oil). Sunflower oil with various levels of oils caused decrease in iodine values (degree of oil unsaturation), this decrease was due to the increase in the predominance of monounsaturated fatty acids of sunflower oil.

Characterization and Identification of SFO by using GC-MS

The oil from Al-Jazeera sunflower was analyzed. Figure. 1. and Table 2. Reflect the profile of fatty acids presence in oil.

The GC-MS analysis of Al-Jazeera sunflower oil revealed some fatty acid with high concentrations, Table.3. Were



Figure 2: GC/MS Chromatogram of Al-Jazeera sunflower oil

R. Time	Area %	IUPAC Name	Formula
12.293	3.07	Phthalic acid, ethyl pentadecyl ester	C25H40O4
12.752	0.38	Cyclooctasiloxane, hexadecamethyl-	C16H48O8
15.875	2.34	Hexadecanoic acid,	C17H34O2
17.344	1.16	1-Nonadecene	C19H38
17.622	4.51	Methyl 10-trans,12-cis-octadecadienoate	C19H34O2
17.666	7.79	9-Octadecenoic acid (Z)-, methyl ester	C19H36O2
19.076	2.19	[1,1'-Bicyclohexyl]-4-carboxylic acid, 4'-pentyl	C24H35FO2
19.130	1.28	Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl	C18H34O2
19.530	1.48	Hexadecanoic acid, 1-(hydroxymethyl)-1,2-ethaned	C35H68O5
20.698	6.22	Oleoyl chloride	C18H33ClO
20.835	1.42	[1,1'-Bicyclohexyl]-4-carboxylic acid, 4'-propyl	C22H31FO2
20.898	1.03	Octadecanoic acid, 2,3-dihydroxypropyl ester	C21H42O4
21.099	14.35	9-Octadecenoic acid, 1,2,3-propanetriyl ester, (E,E,E)	C57H104O6
21.284	3.75	Octadecanoic acid, 2-hydroxy-1,3-propanediyl ester	C39H76O5
22.364	1.09	E,E,Z-1,3,12-Nonadecatriene-5,14-diol	C19H34O2
22.484	4.81	Pentadec-7-ene, 7-bromomethyl-	C16H31Br
22.525	2.22	Octadecanoic acid, 2,3-dihydroxypropyl ester	C21H42O4
22.645	1.25	9-Octadecenoic acid, 1,2,3-propanetriyl ester,	C57H104O6
22.751	2.31	9-Octadecenoic acid, 1,2,3-propanetriyl ester,	C57H104O6
23.466	0.93	D: A-Friedooleanan-7-ol, (7.alpha.)-	C30H52O
23.912	11.53	cis-13-Docosenoyl chloride	C22H41ClO
24.296	16.61	9-Octadecenoic acid, 1,2,3-propanetriyl ester,	C57H104O6
25.540	1.87	9-Undecenal, 2,6,10-trimethyl	C14H26O
25.715	3.08	Heptanoic acid, docosyl ester	C29H58O2
26.500	1.99	Stigmastan-3,5-diene	C29H48
26.856	1.34	18.alphaOlean-3.betaol, acetate	C32H54O2
	100		•

Table 2: GC-MS analysis of the oil extract of Al-Jazeera sunflower

IUPAC Name	Common name	Formula	Content (%)
Hexadecanoic acid	palmitic acid	C15H30O2	7.73
9- Hexadecenoic acid	palmitoleic acid	C16H30O ₂	4.81
Octadecanoic acid,	steric acid	C18H36O ₂	7.00
9-Octadecenoic acid,	Oleic acid	C18H34O ₂	42.31
9-12, Octadecenoic acid,	linoleic acid	C18H32O ₂	19.12

Table 3: The major fatty acids of Al-Jazeera sunflower oil



Figure 3: Fatty acid contents of Aljazeera sunflower oil

The major fatty acids presence in Al-Jazeera sunflower oil were oleic (C18:1), linoleic (C18:2), Stearic (C18:0), palmitoleic (C16:1) and palmitic (C16:0). And acids which together composed about 80.97% of the total fatty acids and 16.96% of other components.

For Al-Jazeera sunflower oil, saturated fatty acids represent 14.73% of the total fatty acids while the unsaturated fatty acid represents 66.24%.

The oil extracted from Al-Jazeera sunflower was analyzed using GC-MS, the result obtained showed high content of fatty acids, rich in oleic acid 42.31%, followed by linoleic acid which was 19.12%. For palmitoleic acid the result showed that the content was 4.81%. Al-Jazeera sunflower oil contain also palmitic acid which was 7.73% and Stearic acid also found in considerable amount which was 7.0%

Change of Physiochemical Characteristics during Storage and Antioxidant Vitamins

Effects of Density (g/cm-¹)

Table 4: Changes in Density (g/cm-1) of SFO during storage (light=L and dark=D) with and without antioxida
vitamin

r											
storage	Density (g/cm ⁻¹)										
storage	storage condition (light vs Dark)										
(week)	SFO ^L	SFO ^L +	SFO ^L +	$SFO^{L}+$	SEODA	SFO ^D +	SFO ^D +	SFO ^D +	value		
(week)	А	Vit. C	Vit. E	Mix vit.	SFU A	vit. C	vit. E	mix vit.			
0	0.9005				0.9005						
0	± 0.001				±0.001						
0	0.9253	0.9206	0.9202	0.9185	0.9193	0.9211	0.9186	0.9184	0.018		
0	± 0.002	±0.001	± 0.001	± 0.001	±.001	± 0.001	± 0.001	± 0.001	0.916		
16	0.9265	0.9247	0.9269	0.9223	0.9296	0.9282	0.9289	0.9308	0.023		
10	±0.01	±0.01	±0.01	±0.003	±0.01	±0.01	±0.01	±0.003	0.925		



Figure 4: effect of light and dark on the Density (g/cm⁻¹) of SFO during storage with and without antioxidant vitamin

Specific gravity is determined and calculated at temperature 20°C as a ratio of mass in air of a given volume of the oil or fat to that of the same volume of at 20°C (Theodore' 1983). It can reveal the extent of adulteration and may be used as a means of acceptance of oils, during the storage time, the density of Aljazeera sunflower oil (SFOL and SFOD) samples stored with and without added vitamins increased slowly and steadily with increase storage time in Table.1. Show the initial density (0.9005 g/cm-1) this value increase with added antioxidants vitamins the density values was calculated after storage time Table.4. And observing in the ranges recommended by the (FAO/WHO, 1993-1994). Standard for edible vegetable oils, probably. This is due to the effect of vitamins on the oils. These results of density are the acceptability of oil quality.

Effects of AV (mg KOH/g of oil)

 Table 5: Changes in AV (mg KOH/g of oil) of Aljazeera sunflower oil during storage (light and dark) with and without antioxidant vitamin

	Acid value (mg KOH/g oil)									
storage time	storage condition (light vs dark)									
(week)	SEOLA	SFO ^L +	SFO ^L +	SFO ^L +	SEODA	SFO ^D +	SFO ^D +	SFO ^D +	value	
	SFU A	Vit. C	Vit. E	Mix vit.	SFU A	Vit. C	Vit. E	Mix vit.		
0	3.3				3.3 ±					
0	±0.02				0.02					
0	3.56	0.42	0.45	0.31	3.5 ±	$0.4 \pm$	0.33 ±	0.29 ±	4	
8	±0.10	±0.07	± 0.07	±0.02	0.13	0.04	0.04	0.10	4	
16	4.80	0.50+0.04	0.59	0.50	3.9 ±	0.49	0.71 ±	0.46 ±		
	±0.12	0.39 ± 0.04	±0.04	± 0.07	0.15	±0.01	0.07	0.02		





During the storage time, the AV of Aljazeera sunflower oil (SFOL and SFOD) samples stored with vitamins decreased slowly and steadily Table.5. The effect of vitamins was high in maintaining the formation of FFA during the eight weeks of mixed vitamins for SFOL and SFOD samples (0.31 \pm 0.02 and 0.29 \pm 0.10) mg KOH/g respectively. And it starts to more capacity as storage time increased. The vitamins E, C and mixed was decreased the AV of Aljazeera sunflower oil (SFOL and SFOD) samples kept at light by compared to SFOL and SFOD control samples stored in light without added vitamin. Similarly, the AV of control samples were presented in this study. Fig.5. Showns higher value than Codex Standard Oils (CODEX-STAN210-1999). The observation of SFOL high values of AV more than SFOD $(4.80 \pm 0.02 \text{ and } 3.9 \pm 0.15 \text{ mg KOH/g of oil})$ respectively indicates a formation of FFAs. Probably, this is due to the effect of light on the oils. The acceptability level of virgin sunflower oils is below 4.0 mg KOH/g. These results of AV decreased oil quality.

At the dark place by $(3.3 \pm 0.02 \text{ and } 3.9 \pm 0.15)$ mg KOH/g from SFOD and stored in similar condition without added vitamins by protecting them against deterioration caused by oxidation which leads to rancidity. This effect shows the antioxidant activity of vitamins in increasing the shelf life of Aljazeera

sunflower oil. The lower AV found, the lower the level of FFA which results in increased oil quality.

The Aljazeera sunflower oil sample kept at dark place show a decrease in the AV content by (0.29 ± 0.10) and 0.46 ± 0.02) mg KOH/g compared to SFOL samples exposed to light. From the samples stored in a dark place, higher change in AV was noted for SFOD than SFOL. This might be related to the presence of high unsaturated fatty acid in SFO, especially oleic acid (18: 1), which is susceptible to oxidation, hydrolysis, and thermal degradation. It may be postulated that light absorption was greater in SFOL as compared to darkness. This result is in line with works reported on sunflower and rapeseed oil under dark storage by (Abramovic and Abram, 2005).

The Vitamins functions as an antioxidant by serving as free-radical terminators and scavenging singlet oxygen molecules. The ascorbic acid and α -tocopherol concentration is an important factor that effects antioxidant activity in oils. Studies in purified TAGs obtained from Aljazeera SFO showed that antioxidant activity α -tocopherol was greater at concentrations (200 ppm) and it loses efficacy at higher concentrations due to its participation inside reactions. Figure 5. Shows the AV changes of oils stored under light and dark after the addition of 200 mg/kg vitamin E and C.

Effects of PV (meq O₂/kg oil)

Table 6: Changes in PV (meq O2/kg oil) of SFO during storage (light and dark) with and without antioxidant vitamin

	Peroxide value (meq O ₂ /kg oil)								
storage time	storage condition(light vs Dark)								
(week)	SEOLA	SFO ^L +	SFO ^L +	SFO ^L +	SEOD A	SFO ^D +	SFO ^D +	SFO ^D +	value
	SFU A	Vit. C	Vit. E	Mix vit.	3FU +A	Vit. C	Vit. E	Mix vit.	
0	$7.28 \pm$				$7.28 \pm$				
0	(0.04)				(0.04)				
8	$7.28 \pm$	3.4 ±	3.48 ±	2.73 ±	$4.97 \pm$	3.48 ±	2.59 ±	2.14 ±	15
	(0.40)	(0.13)	(0.20)	0.18)	(0.27)	(0.20)	(0.12)	(0.04)	15
16	9.69 ±	7.53 ±	$6.63 \pm$	$5.66 \pm$	$7.68 \pm$	5.64 ±	6.29 ±	4.19 ±	
	(0.83)	(0.89)	(0.33)	(0.50)	(0.83)	(0.89)	(0.33)	(0.77)	



Figure 6: effect of light and dark on the PV of SFO during storage with and without antioxidant vitamin

At the beginning of the experiment, the PV of SFOL and SFOD was (7.28 \pm 0.04, 9.69 \pm 0.83 and 7.68 \pm 0.83, 7.68 \pm 0.83) meq O₂/kg. This value falls in the range considered as satisfactory and in agreement with the maximum Codex standard PV (15 meg O2/Kg) (Atta et al., 2008). For virgin vegetable oil. The PV of oil samples stored for 16 weeks registered a progressive increase with the increment of storage period. From Table 6 it was observed that changes in PV of SFOL and SFOD stored under different conditions with and without added vitamins were significant (p<0.05). At the end of 16 weeks of storage, the Table.6. Shows the PV of SFOL and SFOD samples stored at light accelerates oxidation more than dark. The change in PV of Al-Jazeera SFO kept at light was significantly (p<0.05) higher than the PV value of Al-Jazeera SFO stored at dark. Lower change in PV of Al-Jazeera compared to SFOD dark vitamin during light storage was probably due to higher content of saturated palmitic acid, which is less prone to oxidation than unsaturated fatty acids, linoleic acid and Oleic acid. (Paul, et al., 1992)

The higher PV of control SFOL and SFOD is mostly because Al-Jazeera SFO has an appreciable **Table 7: Changes in IV** (gI₂/100g oil) of SFO during storage (light and dark) with and without antioxidant vitamin

amount of unsaturated FA to fix oxygen and easily oxidized. These findings are similar to the work of Huang *et al.* 1981) who reported that high PUFAs, especially linoleic acid (18:2), are prone to oxidation, hydrolysis, and thermal degradation.

The absence of light lowered the PV of SFOD by (7.28 and 2.14) meq O2/kg the first eight weeks storage when compared to SFOL stored in light, this finding shows the oxidation process during periods of storage was affected by light. Figure 6. Shows the PV changes of oils stored under light and dark after the addition of 200 mg/kg vitamin E and C.

A lot of literature states that faster oxidation occurs due to exposure to light (Khan and Shahidi 2002). The absence of light minimized the hydroperoxide formation and also synergistically supported by minor components found in oils which acts as an antioxidant in the dark (⁶Khan and Shahidi, 2000). Such decreases in PVs had been reported by Neff *et al.*, 1994).

Effects of Iodine Value (IV)

Storage		standard							
time (week)	SFO ^L A	SFO ^L + Vit. C	SFO ^L + Vit. E	SFO ^L + Mix vit.	SFO ^D A	SFO ^D + Vit. C	SFO ^D + Vit. E	SFO ^D + Mix vit.	value
0	134.5 ± 0.71				134.5± 0.71				110
8	98.53 ± 0.76	106.8 ±4.76	106.76 ±7.03	107.44 ± 7.17	97.51 ±1.11	107.46 ±6.78	101.85 ±7.35	101.39 ±8.09	143
16	97.1 ± 0.96	98.51 ±1.24	96.48± 1.41	98.11± 0.16	98.43 ±0.96	97.08 ±1.24	99.13 ±1.41	98.48 ±0.16	





Changes in Iodine Value (IV) During Storage

For Sunflower Oil, the high iodine value portrays that it is rich in unsaturated fatty acid which implies that it will have short oxidative storage stability because according to Perkins the oxidative and chemical changes in oils during storage are characterized by increase in FFA content and a decrease in the total unsaturation of oils (Perkins, 1992). Iodine value is a measure of the degree of unsaturation or double bonds among the fatty acid present in the oil therefore it does not tell precisely the fatty acids composition of any oil. Iodine value or number is useful as a guide to check adulteration of oil and also as a process control of oil. Free Fatty Acid values was lower for the oil as compared to the value recommended by FAO/WHO which may be attributed to the variation in variety, and climatic conditions which is evident in the iodine value. Fig.7. the effect of vitamin E and C, was high in maintaining the formation of FFA during the Sixteen weeks.

The different results for storage conditions of SFO significantly differed in iodine values due to the insignificant variation in their fatty acid composition Table.7. Iodine values showed significant and positive correlations with FAs and other polyunsaturated content. Data indicated that the SFOL and SFOD had the lower in iodine values. The iodine values were in the ranges recommended by the (Codex Standard, 2003). The PUFA ratios as well as iodine values were indicative of unsaturation levels and as a result,

The oil has a tendency to undergo autoxidation (Farhoosh, *et al.*, 2008). Decreased levels of unsaturation linoleic acid will result in increased levels of oxidative stability. Therefore the high oleic sunflower oil with their lower levels of unsaturation should be more resistant to oxidation than the high linoleic acid sunflower oil.

CONCLUSION

Oxidative stability is an important indicator of oil quality and shelf-life. The oxidative changes during storage and domestic use make oils unsuitable for consumption. The present investigation is an overview of changes in acid value, peroxide value and Iodine Value of sunflower oil from 16 weeks of storage. The results observed during the study show that prolonged storage of sunflower oil at ambient (25-33) °C temperatures can lead to oxidative deterioration of oil samples. Initially, oil samples show acceptable acid value from the recommended value and significantly increased during storage. Higher change in acid value observed for a sample stored in light than dark. Also, additions of vitamins decreased the formation of free fatty acids in sunflower oil. The initial peroxide value of both oil samples is in agreement with Codex standard guideline value. The peroxide value significantly increased during storage. Higher peroxide value was observed for samples stored in light than samples stored in dark. Also, the addition of vitamins decreased the change in the peroxide value of sunflower oil. The added vitamins prohibited the formation of hydroperoxide more in the dark than light storage.

The high content of primary oxidation products and the presence of FFAs might profoundly lower the oxidative stability of the oil. Even if the oxidation stability of SFOL was found to be higher than oxidative stability of sunflower oil in the dark from the acid value, peroxide value obtained, both SFOL and SFOD are acceptable during storage with added vitamins as antioxidants. In general, the results of the present study show that light acts as a major catalyst in accelerating the development of rancidity in oils. However; the addition of vitamins to oil can increase the oxidation stability of oils during storage. Storing in dark (packaging with material protect light) and supporting with antioxidant vitamins is the best way to maintain the quality of oils during storage and domestic uses.

RECOMMENDATION

The investigations presented in this study suggest:

- Better packing and storage conditions can lead to an improvement in the oxidative stability of vegetable oils and other related products containing fats and oils
- Lipid oxidation products make the oil unfit for human health; therefore, to minimize the oxidation phenomenon, some antioxidants should be added to increase the storage and shelf life of oils and oil products
- Producers, shopkeepers, and users should store oils and oil products in dark places protected from light
- The government or any responsible body should follow that local oil producers have put the production and expiry dates of domestically produced oils to safeguard the health of people

REFERENCES

- Abramovič H. and Abram V., 2005. Physico-chemical properties, composition and oxidative stability of Camelina sativa oil. Food Technol Biotechnol., 43:63-70.
- Alimentarius C., 1999. Codex standard for named vegetable oils. Codex stan, **210**:1-3.

- A. C., 2005. Official methods of analysis of the association of official analytical chemists, 18th Edn., (Edited by W. Howitz). Washington, DC: AOAC.
- A. O. A. C., 2000. Official methods of analysis 17th Edition, Association of Official Agric Chem. Washington, D.C: 1970.
- A.O.C.S., 2003. Official Method Cd 8-53. American Oil Chemists Society, Champaign, IL.
- Atta S., Ahmad T., Gul S., Nagra S.A., Luthfullah G. and Khan M., 2008. Effect of gamma irradiation on the nutritional quality of ready to eat fast food. J Chem Soc Pak., 30:879-883.
- Codex Standard, 2003. Codex standard for named vegetable oils. Codex Stan 210-1999. pp: 1-16.
- Farhoosh R., Niazmand R., Rezaei M. and Sarabi M., 2008. Kinetic parameter determination of vegetable oil oxidation under rancimat test conditions. Eur. J. Lipid Sci. Technol., **110**(6): 587-592.
- Frankel E.N., 2005). Lipid Oxidation, Bridgwater, England, the Oily Press.
- Gizachew Z., 2020. Influence of Storage Condition and Added Vitamin E on Oxidation Stability of Edible Sunflower and Nigger Seed Oils Produced and Sold around Bahir Dar City, Ethiopia. J. Food Process Technol., **2**(11):821.
- Huang A.S., Hsieh O.A., Huang C.L. and Chang S.S., 1981. A comparison of the stability of sunflower oil and corn oil. J Am Oil Chem Soc., **58**:997-1001.
- I.O.O.C., 2009. International olive oil council. International trade standard Appling to olive oils and olive- pomace oils. COI/T.15/NC 3/Rev. 2.
- Khan M.A. and Shahidi F., 2002. Photooxidative stability of stripped and nonstripped borage and evening primrose oils and their emulsions in water. Food Chem., **79**:47-53.
- Khan M.A. and Shahidi F., 2000. Tocopherols and phospholipids enhance the oxidative stability of borage and evening primrose triacylglycerols. J. Food lipids., 7:143-150.
- Khan S., Choudhary S., Pandey A., Khan M.K. and Thomas G., 2015. Sunflower oil: Efficient oil source for human consumption. Emergent Life Sciences Research, **1**: 1-3.

- Fu M., Xu S., Hui P., Qiang Z., Jing Y., Yue S., Ho C.-T., Huimei C. and Ruyan H., 2019. Identification of rancidity markers in roasted sunflower seeds produced from raw materials stored for different periods of time. J.L.W.T., 118: 108721.
- Monotti M., 2004. Growing non-food sunflower in dry land conditions. Ital. J. Agron., **8**(1): 3-8.
- Neff W.E., Mounts T.L., Rinsch W.M., Konishi H. and ElAgaimy M.A., 1994. Oxidative stability of purified canola oil triacylglycerols with altered fatty acid compositions as affected by triacylglycerol composition and structure. J. Am. Oil Chem. Soc., **71**:1101-1109.
- Okpuzor J., Okochi V.I., Ogbunugafor H.A., Ogbonnia S., Fagbayi T. and Obidiegwu C., 2009. Estimation of cholesterol level in different brands of vegetable oils. Pak. J. Nutr., 8:57-62.
- Paul J.N., Manny N., Baryan B.G., Rosemary M., Peter M.C. and Mavis A., 1992. Plasma lipoprotein lipid and lipid changes with substitution of elaidic acid for oleic acid in the diet. Am. J. Clin. Nutr., 55:46-50.
- Perkins E.G., 1992. Effect of lipid oxidation on oil and food quality in deep frying: Angelo AJS (ed) Lipid oxidation in food. ACS Symposium ACS publications American Chemical Society Washington DC., 18(500):310-321
- Prevention of Food Adulteration Act-1954, Commercial Law Publishers Pvt. Ltd. Delhi, India, 2005.
- Sarkar S., Mukherjee A., Swarnakar S. and Das N., 2016. Nanocapsulated ascorbic acid in combating cerebral ischemia reperfusion-induced oxidative injury in rat brain. Current Alzheimer Research., 13:1363.
- Theodore A.G., 1983. Food Science. 3rd Edition AVI Publishers, West Port.
- Traber M.G., Shils M.E., Shike M., Ross A.C., Caballero
 B. and Cousins R., 2006. eds. Vitamin E.
 Modern Nutrition in Health and Disease. 10th
 edition Baltimore, MD: Lippincott Williams & Wilkins, 396-411.
- Traber M.G., 2007. Vitamin E regulatory mechanisms. Annu. Rev. Nutr., **27**:347-62.
- W.H.O. Fats and oils in human nutrition Report of a Joint FAO/WHO Expert Consultation Committee, Rome, Italy, World Health Organization, Geneva, (1993-1994).