SYNERGETIC EFFICACY OF THREE GYMNOSPERMS AGAINST Klebsiella pneumoniea

RAVIKANT SINGH^a, ANUPAM DIKSHIT^b AND ROHIT K. MISHRA^{c1}

^{ac}Department of Biotechnology, Swami Vivekanand University, Sagar, M.P., India
^bDepartment of Botany, University of Allahabad, Allahabad, U.P., India

ABSTRACT

The present investigation were focused on the synergetic efficacy of the essential oil of the three gymnospermous foliages *i.e., Pinus roxburghii* Sarg., *Taxodium distichum* L. and *Thuja occidentalis* L., against *Klebsiella pneumoniea* (MTCC-4032). The oils were extracted from the needles and foliages of aforementioned plant species using hydro-distillation method. The antibacterial activity of the extracted essential oils was evaluated against *Klebsiella pneumoniea* (MTCC- 4032) using broth micro-dilution method recommended by Clinical Laboratory Standards Institute (CLSI). The Inhibition Concentration *i.e.* IC₅₀ and Minimum Inhibition concentrations (MIC) using SpectramaxPlus³⁸⁴, of Molecular Devices Corporation, USA were recorded while Streptomycin as standard was taken. The IC₅₀ value of *P. roxburghii*, *T. distichum* and *T. occidentalis* were showed 0.278, 0.093 and 0.161 mg/ml respectively. The *T. distichum* was found most effective with their MIC 0.277 mg/ml while *P. roxburghii* found least effective with their MIC 0.616 mg/ml against *K. pneumoniea*. Hence, essential oil from foliages and needles of gymnosperms exhibit great potential for the development of eco-friendly, non-toxic, cost effective anti-bacterial formulations after undergoing detailed investigation which is in progress.

KEYWORDS: Gymnosperms, Essential Oil, Synergetic Efficacy, Broth Micro-dilution

P. roxburghii Sarg. is also known as chir pine, belongs to family Pinaceae, is a native of Himalayas and distributed throughout India, Nepal, Bhutan and Pakistan. It is widely distributed throughout Himalayan range of India.P. roxburghii is a large tree attaining up to 30-60 m in height with a trunk diameter reaching up to 2 m (figure 1). Ovoid and conic cones of P. roxburghiiusually open up to 18-20 cm to release the seeds (Press et al., 2000). P. roxburghii oil has been traditionally used to treat cuts, wounds, boils and blisters (Wu and Raven, 1999). In addition, phytochemical screening of Pinus needles and stems have pound abundant amounts of vitamin C, tannins, and alkaloids while the stem has been primarily used as a source of turpentine oil (Gewali, 2008) (Vallejo et al., 1994) (Asta et al., 2006). Previous research on microbes suggests that the essential oil on P. roxburghii has shown significant anti-fungal and anti-bacterial activity (Hassan and Amjid, 2009) while alcoholic extract of the needle, stem, and cones are reported to exhibit strong anti-bacterial activity.

Taxodium distichum. (L.) L. C. is commonly known as bald cypress, is an unusual and interesting tree, belongs to Taxodiaceae. It may grow upto 25 m in height and over 300 cm in diameter (figure 2). The leaves are small, 5–15 mm long, green to yellow-green and appearing two-ranked. The fruits are cones and are composed of scales forming a woody, brown sphere with rough surface

1.5 to 4 cm in diameter. Taxodium distichum (L.) has three extant taxa ranging from the eastern United States through Mexico to Guatemala (Adams, 2001). The trees are used for their wood because heartwood is extremely rot and termite resistant (Tantawy et al., 1999). Leaves and cones are rich in essential oils and used traditionally to treat gastro-intestinal, skin, respiratory, inflammation, and infections (Flamini et al., 2000) (Geiger and de Groot-Pfleiderer, 1979).Flavonoids and diterpenoids are the main secondary metabolites (Adams, 2001). T. distichum trees can grow on rivers, lake margins, swamps, wet poorly drained habitats and are tolerant to various soil conditions and air pollution (Denny, 2007). In many countries these are widely used for landscape. The heartwood of bald cypress is used for building materials, and has been reported to resist the attacks of the subterranean termite (Kusumoto et al., 2010).



Figure 1: P. roxburghii



Figure 2: T. distichum



Figure 3: T. occidentalis

Thuja occidentalis L. is the native to Eastern Canada and other regions on United State; widely cultivated as an ornamental plant (figure 3). *T. occidentalis* has been used to treat bronchial catarrh, psoriasis, rheumatism and uterine carcinomas (Peng and Wang, 2008). The essential oil of the plant has been used for disinfectants, insecticides, room sprays, and soft soaps. Cedar leaf oil can be obtained by steam distillation or hydro-distillation of the foliage and is used for the production of perfumes, insecticides, soaps and deodorants (Kamden and Hanover, 1993) (Duke, 1985). The essential oil is an active ingredient in the production of cough suppressants, perfumes and soaps, while many cultivars are grown for ornamental purposes (FAO, 1995).

MATERIALS AND METHODS

Extraction of Essential Oil

The plant materials of *P. roxburghii*, *T. distichum*, and *T. occidentalis* were collected from Roxburgh Garden, Department of Botany, University of Allahabad, in the month of October. Plant were identified at Department of Botany, University of Allahabad. Leaves (needles, foliages) and branchlets were crushed and hydrolyzed using a Clevenger type Apparatus for 4 hours (figure 4). Secondary metabolites of *T. distichum* (bald cypress) appears as dark yellow, yellow coloured oil were extracted from *T. occidentalis* followed by *P. roxburghii* (chir pine) i.e., pale yellow (figure 5). Oil content was stored at 4°C until analysis (Isiaka *et al.*, 2007).



Figure 4: Clevenger Type Apparatus.



Figure 5: Extracted Oils

Preparation of 0.5 McFarland Solution and Saline Media

Standard method was slightly modified for our study. Dissolve 2.04 ml of H_2SO_4 in double distilled water (DDW). Now add 1% BaCl₂ to the freshly prepared solution (McFarland, 1907). 0.5 McFarland solution is ready. To prepare saline media, dissolve 1 gm of NaCl into 100 ml of DDW (Singh *et al.*, 2018). Take the O.D. of this saline media.

Preparation of Mueller-Hinton Broth (MHB)

Take 250 ml of DDW in a beaker. Add 5.25 gms of MHB powder. Shake well and boil up to 100 °C. Close the mouth with cotton plug. Place the solution inside autoclave. After this, MHB is ready to use.

Preparation of Inocula

Saline media was taken in a culture tube. Add bacterium in the same tube. Final volume in the culture tube is 1000μ L. Now take 500μ L of this solution and add it into another culture tube containing MHB, so that the final volume becomes 20 ml. Now inocula will be ready for use.

Antibacterial Screening

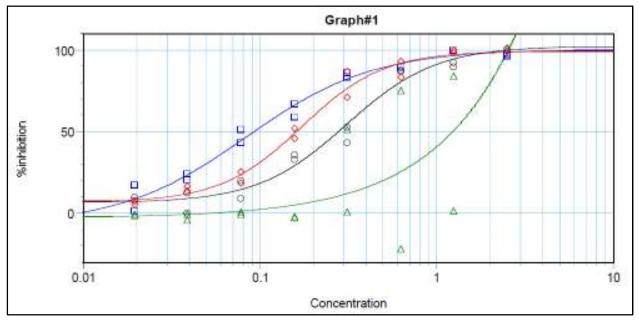
Essential oils were screened for antibacterial activity against *K. pneumoniea*. Minimum Inhibitory Concentrations (MIC) were determined using Broth Micro-

dilution method recommended by Clinical Laboratory Standard Institute (CLSI). 96 well plate was used for microdilutions. Column-1 contains 190µL and 10 µL of after the formaldehyde (added completion of microdilution). Column-2 contains 200µL of MHB. Column-3 is the drug control. Row A and B of column-3 contains streptomycin. Row C and D containsPinus oil. Row E and F contains Taxodium oil whereas row G and H contains Thuja oil. 100 µL of broth were added from column-4 to column-12. In column-4, we add drugs in each row one by one as described previously for column-3. Now dilute the drugs horizontally from column-4 to column-11. Now add 100µL inocula to each well from column-4 to column-11. Final volume of each well were 200µL.The extract solutions over horizontally diluted 1:1 in MHB in a 96 well plates were incubated at 37 °Cfor 24 hours (Satyal et al., 2012) (Singh et al., 2018). The final minimum inhibitory concentration and it was determined as the lowest concentration without turbidity. Streptomycin used as positive control. Formaldehyde was used as a negative control.

RESULTS

Percent yield: % yield = weight of oil / weight of sample x 100.

P.roxburghii = 0.100 %, *T. distichum* = 0.260% and *T. occidentalis* = 0.350 %.



Graph 1: Graph Showing Percent Inhibition against Concentration of Different Drugs.

```
IC50 Determinations
IC-50 Of the four drugs is as follows :-
IC50-1 = 1.232
IC50-2 = 0.278
IC50-3 = 0.093
IC50-4 = 0.161
MIC of the four drugs is as follows :-
MIC-1 = 2.008
MIC-2 = 0.616
MIC-3 = 0.277
MIC-4 = 0.350
```

Figure 6: IC50 and MIC values obtained from Spectra Max Plus 384.

The results were recorded in terms oil Inhibition (IC50) and Minimum Inhibition Concentrations Concentrations (MICs) via SpectramaxPlus384, Molecular Devices Corporation, USA. IC50 value of P. roxburghii, T. distichum and T. occidentalis were showed 0.278, 0.093 and 0.161 mg/ml respectively (Figure 6). The minimum inhibition concentrations (MIC) of P. roxburghii, T. distichum and Thuja occidentalis were recorded 0.616, 0.277 and 0.350 mg/ml respectively (Figure 6). T. distichum was found to be most effective with their MIC 0.277 mg/m1 whereas P. roxburghii was found to be least effective with their MIC 0.616 mg/m1 against K. pneumoniea.

CONCLUSION

It can be concluded from the present study that all the three Gymnospermous essential oil have some activity against K. pneumoniea. Taxodium oil shows remarkable efficiency over Pinus oil and Thuja oil against bacteria. Taxodium oil shows great efficiency against K. pneumoniea and other microbes (Von Rudloff, 1975). The components of essential oil of P. roxburghii needles are highly active against microbes (Singh et al., 2018). As this oil significantly inhibited the growth of certain bacteria and fungi tested. The main oil component of P. roxburghii essential oil are monoterpene and sequiterpene hydrocarbons and their derivatives. These derivatives act as antibacterial and antifungal substance, the most well-known of which being terpenes and phenolics in general (Gulten et al., 2012). The essential oil from the leaves and cones of baldcypress trees grown exhibited potent antimicrobial activities against bacteria (Tantawy *et al.*, 1999).So, essential oils of these gymnosperms exhibit great potential eco-friendly, non-toxic, cost-efficient and antibacterial herbal formulations (Singh *et al.*, 2018).

ACKNOWLEDGEMENTS

I would like to give special thanks to Prof. D.K. Chauhan (HOD) Department of Botany, University Of Allahabad, for providing me laboratory facilities; Mr. Saket Jha and Mr. Sharad Kumar Tripathi for their valuable suggestions.

REFERENCES

- Adams R.P., 2001. Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry. Carol Stream, IL, Allurred Publishing Corporation, pp. 1–456.
- Adams R.P., 2001. Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry. Carol Stream, IL, Allurred Publishing Corporation, pp. 1–456. (Google Scholar).
- Asta J., Jurgita S., Aida S. and Eugenija K., 2006. Characteristics of essential oil composition in the needles of young scots pine (*Pinus sylvestris* L.) stands growing along and ariel ammonia gradient. Chemija, 17:67–73.

- Denny G.C., 2007. Evaluation of selected provenances of Taxodium distichum for drought, alkalinity and salinity tolerance. PhD Thesis, A&M University: Texas.
- Duke J.A., 1985. Handbook of Medicinal Herbs; CRC Press, Inc.: Boca Raton, FL, USA.
- Flamini G., Luigi C. and Morelli I., 2000. Investigation of the essential oil of feminine cones, leaves and branches of *Taxodium distichum*. from Italy. J. Essent Oil Res., **12**: 310–312.
- FAO (Food and Agriculture Organization of the United Nations), 1995. Non-Wood Forest Products from Conifers. Chapter 7-Essential Oils; FAO: Rome, Italy, 12: 86.
- Geiger H. and de Groot-Pfleiderer W., 1979. Die flavonund flavonolglykoside von *Taxodium distichum*.. Phytochemistry, **18**: 1709–1710
- Gewali M.B., 2008. Institute of Natural Medicine. Japan: University of Toyama. Aspects of Traditional Medicine in Nepal; pp. 19–20.
- Gulten T.G., Branden A.N., Sahika A.G. and Mehmat K., 2012. "Antimicrobial activity of oregano oil on Iceburg lettuce with different attachment conditions." J. Food Sci., **77**(7): 412-415.
- Hassan A. and Amjid I., 2009. Gas chromatography-mass spectrometric studies of essential oil of *Pinus roxburghaii* stems and their antibacterial and antifungal activities. J. Med. Plant Res., **3**:670–3.
- Isiaka A. Ogunwande, Nureni O. Olawore, Oluranti O. Ogunmola, Tameka M. Walker, Jennifer M. Schmidt and William N. Setzer, 2007. Cytotoxic Effects of Taxodium distichum. Oils, Pharmaceutical Biology, 45(2): 106-110.
- Kamden P.D. and Hanover J.W., 1993. "Inter-Tree variation of essential oil composition of Thuja occidentalis L." J. Essent. Oil Res., **5**: 279–282.
- Kusumoto N., Ashitani T., Murayama T., Ogiyama K. and Takahashi K., 2010. Antifungal abietane-type diterpenes from the cones of Taxodium distichum Rich. J. Chem. Ecol., **36**(12): 1381- 1386.

- McFarland J., 1907. Nephelometer: an instrument for media used for estimating the number of bacteria in suspensions used for calculating the opsonic index and for vaccines. J. Am. Med. Assoc., 14:1176-8.
- Peng D. and Wang X.-Q., 2008. Reticulate evolution in Thuja inferred from multiple gene sequences: Implications for the study of biogeographical disjunction between easren Asia and North America. Mol. Phylogenet. Evol., 47: 1190–1202.
- Press J.R., Shrestha K.K. and Sutton D.A., 2000. Annotated Checklist of the Flowering Plants of Nepal. The Natural History Museum.
- Singh R.K., Singh R., Singh P., Jha S., Pathak A., Shukla S.K., Dikshit A. and Mishra R.K., 2018. Synergistic effect of Essential oils of three Gymnosperms against *Vibrio cholera*. Inter. Jour. Res. Appl. Sci. Eng. Tech. (IJRASET). 6(3): 2321-9653.
- Singh R.K., Pathak A., Dikshit A. and Mishra R.K., 2018. Comparison of biological activities of essential oil of three Gymnosperms against *Salmonella typhimurium*", Intl. Jour. Crea. Res. Thou. (IJCRT). ISSN: 2320-2882. 6(1): 1420-1426.
- Singh R.K., Pathak A., Tripathi S.K., Dikshit A. and Mishra R.K., 2018. A Reassessment on Essential Oils. Intl. Jour. Sci. Eng. Develop. Res. (www.ijrti.org), ISSN: 2455-2631. 3(6): 177 - 187.
- Singh R.K., Pathak A., Dikshit A. and Mishra R.K., 2018. Antibacterial evaluation of essential oils of Juniperus communis, Pinus roxburghii and Thuja occidentalis against Escherichia coli. Biochem. Cell. Arch., 18(1): 241-244.
- Singh R.K., Jha S., Pandey A., Dikshit A. and Mishra R.K.,
 2018. "Comparison of Antibacterial Activities of essential oils of Juniperus communis L., Pinus roxburghii Sarg. and Taxodium distichum L. against Klebsiella pneumoniea", International Journal of Emerging Technologies and Innovative Research (www.jetir.org), ISSN: 2349-5162, 5(8): 661-669. http://www.jetir.org/papers/JETIR 1808400.pdf.

- Satyal P., Paudel P., Lamichhane B. and Setzer W.N., 2012. Volatile constituents and biological activities of the leaf essential of *Jasminum mesnyi* growing in Nepal. J. Chem. Pharm. Res., 4:437–9.
- Tantawy M.E., Sakhawy F.S., Sow M. and Ross S.A., 1999. Chemical Composition and Biological Activity of the Essential Oil of the Fruit of *Taxodium distichurn* L. Rich Growing in Egypt. J. Essent. Oil Res., 11: 386-392.
- Tantawy M.E., Sakhawy F.S., Sohly M.A. and Ross S.A., 1999. Chemical composition and biological activity of the essential oil of the fruit of *Taxodium distichum*. L. growing in Egypt. J.

Essent Oil Res., **11**: 386–392 (and references cited therein).

- Von Rudloff E., 1975. Volatile Leaf Oil Analysis in Chemosystematic Studies of North American Conifers. Biochem. Sys. Ecol., 2: 131–167.
- Vallejo M.C.N., Evandro A. and Sergio A.L.M., 1994.
 Volatile wood oils of the Brazilian *Pinus* caribaea var. hondurensis and Spanish *Pinus* pinaster var. mediterranea. J. Braz. Chem. Soc., 5:107–112.
 - Wu Z. and Raven P.H., 1999. Flora of China. Vol. 4. Beijing Science Press; 1999.