

BLA-NDM-1 IN CLINICAL ISOLATES OF *Acinetobacter baumannii* FROM NORTH INDIA

NIDHI PAL^a, R. SUJATHA^b AND ANIL KUMAR^{1c}

^aDepartment of Microbiology, Rama Medical College and Research Centre, Mandhana Kanpur (UP) India.

^bProfessor and Head, Department of Microbiology, Rama Medical College Hospital and Research Centre, Mandhana, Kanpur (UP) India

^cAssistant Professor, Central Research Laboratory, Rama Medical College Hospital and Research Centre, Mandhana, Kanpur, (UP) India

ABSTRACT

The growing incidence of the resistance to carbapenems due to NDM-1 in *Acinetobacter baumannii* is of major concern throughout the world. In this study 200, test cases were studied from October 2015 to January 2017 from six different hospitals of Kanpur. *A.baumannii* were isolated and identified by standard procedure. *A.baumannii* were confirmed genotypically by PCR using OXA 51 primer. All the isolated *A.baumannii* were screened for antimicrobial resistant pattern according to CLSI guideline. The screened isolates were further phenotypically studied for carbapenemase production by the MHT and CDDT. All the screened isolates were also subjected to PCR detection of the blaNDM-1 gene. From 200 patients admitted to ICU of different tertiary care hospitals and total 34 *A.baumannii* was isolated. The prevalence of *A.baumannii* infection in Kanpur region was 10% (20 out of 200 cases) Table 1. The rate of *Acinetobacter* infection was 9.18 per 1000 ICU days. Antimicrobial resistance pattern of isolated *A.baumannii* was also recorded and it found that only Polymyxins and Tigecycline showed 100% sensitivity. 27 strains were carbapenem resistant. Phenotypically only 18 strains were found to produce carbapenemases by CDDT and 11 strains were MHT positive. Among 34 isolated *A. baumannii*, 11 samples were found positive for bla_{NDM-1}.

KEY WORDS: bla_{NDM-1}, Polymyxins, carbapenem, *A. baumannii*, nosocomial infections

NDM-1 is a novel type of metallo- β -lactamase that hydrolyzes all the β -lactam antibiotics. The gene that encodes for NDM-1 is called blaNDM-1 and is located on a transmissible plasmid and its association with other resistant determinants leads to the extensive drug resistance which is exhibited by a majority of the NDM-1 producing microorganism, leaving only a few therapeutic options (Young et al., 2009 and Kumarawamy et al., 2010)

Therefore, the NDM-1 producing organisms are now being recognized as the world's newest superbugs. In general, the term "superbugs" is a colloquial reference to a bacterium that carries resistance genes for many antibiotics (Salcido, 2010).

Acinetobacter baumannii, has emerged as an important opportunistic Gram-negative bacteria in health care institutions globally, as it resists desiccation, is hard to eradicate and has numerous intrinsic and acquired mechanisms of drug resistance.

However, the data on the prevalence of NDM-1 producing *A. baumannii* isolates in Indian hospitals is limited. We therefore designed the Present study to evaluate the occurrence of carbapenem resistant *A. baumannii* and blaNDM-1 in the *A. baumannii* isolates in Kanpur.

MATERIALS AND METHODS

The Bacterial Isolates

A total of 34 non-duplicated *A. baumannii* isolates were isolated from various clinical samples such as Endotracheal aspirate, Endotracheal tip, sputum, blood, central line tip, urine and urinary catheters at different hospitals of Kanpur (UP). The samples were obtained from ICU admitted patients more than 48 hours between October 2015 and January 2017. The standard microbiological techniques were used for the isolation and the identification of the isolates has been mentioned in our study (Pal et al., 2017). All the isolates were stored in 10% glycerol-supplemented Brain heart infusion broth at -20^oC. This study was carried out with the consent of the institutional ethics committee.

¹Corresponding Author

Antimicrobial Susceptibility Testing

The Kirby-Bauer disc diffusion method was performed to determine the susceptibilities of the different antibiotics and the results were interpreted as per the Clinical and Laboratory Standards Institute (CLSI) guidelines (CLSI, 2016). All the antibiotic discs and the media were procured from Hi-media, Mumbai, India. The *E. coli* ATCC 25922 and the *Pseudomonas aeruginosa* ATCC 27853 strains were used for quality control.

Screening for the Carbapenemase Production

By disc diffusion, all the isolates resistant to imipenem and meropenem (diameter of zones of inhibition, ≤ 21 mm) were screened for the production of carbapenemase according to the CLSI guidelines

Phenotypic Detection of the Carbapenemase Production

The phenotypic detection of the carbapenemase production was performed by the Modified Hodge Test (MHT) by using a imipenem disc (10 μ g). The detection of metallo- β -lactamase production was also performed by the Combined-Disc Diffusion Test by using two imipenem discs (10 μ g), one containing 10 μ l of 0.1 M EDTA, which were placed 20 mm apart on a Mueller-Hinton agar plate. An increase in the zone diameter of >5 mm around the imipenem-EDTA disc as compared to that of the imipenem disc alone was considered as positive for metallo- β -lactamase production.

Molecular detection of the bla_{NDM-1} genes

DNA was extracted from all the screening positive isolates by using the DNA extraction Kit (Qiagen, Germany) as per the manufacturer's protocol.

PCR reaction

PCR was carried out in 20 μ l reaction volume with 10 μ l master mix (Takara), 5 μ l nuclease free water, 1 μ l forward and reverse primer [NDM-Fm (5'-CTGAGCACCGCATTAGCC-3') and NDM-Rm (5'-GGCCGTATGAGTGATTGC-3') (GCC biotech, Calcutta)] each and 3 μ l DNA template. Conditions for PCR were as below, initial

denaturation 94 °C for 5 min, number of cycles were 34 in each cycle denaturation was set at 94 °C for 30 sec, annealing at 50 °C for 30 sec and then 72°C for 1 min for extension, followed by a final extension at 72°C for 5 min. After amplification, amplified DNA was identified by 1% agarose gel electrophoresis. Loading and electrophoresis of amplified PCR product and visualization of the band on agarose gel were performed as mentioned before.

The PCR product of the bla_{NDM-1} gene was sent for sequencing to Chromus biotech Pvt. Ltd., Bangalore, India.

RESULTS

In this study 200, test cases were studied from October 2015 to January 2017 from six different hospitals of Kanpur. From 200 patients admitted to different ICU, total 315 samples were collected and microbiologically processed.

In this study total of 200 patients were observed for nosocomial infections, admitted to ICU of different tertiary care hospitals and total 34 *Acinetobacter baumannii* was isolated. These 34 isolated *A. baumannii* also gave the positive result in PCR with OXA 51 primer.

On the basis of microbiological and clinical correlation among 34 isolated *A. baumannii*, 20 were contributed to the infection. Hence, the prevalence of *A. baumannii* infection in Kanpur region was 10% (20 out of 200 cases) Table 1. The rate of *Acinetobacter* infection was 9.18 per 1000 ICU days. Antimicrobial resistance pattern of isolated *A. baumannii* were shown in table 2. Only Polymyxins and Tigecycline showed 100% sensitivity. 27 strains were carbapenem resistant. Phenotypically only 18 strains were found to produce carbapenemases by CCDT [Table 3]. While only 11 strains were MHT positive. [Table 3] Sensitivity and specificity of tests have mentioned in table 4. After the analysis of phenotypically all the isolated *A. baumannii* strains were subjected to bla_{NDM} detection. Among 34 isolated *A. baumannii*, 11 samples were found positive for bla_{NDM-1} [Fig 1].

Table 1: Distribution of *A. baumannii* associated with infection and colonization

Infection/colonizer	No. of <i>A. baumannii</i>
Associated with infection	20
Associated with colonizer	14

Table 2: Antibiotic resistant pattern of *A. baumannii*

S.N.	Antibiotics	Antibiotics	No. of Strains	Percent
1.	Piperacillin(100µg)	PI	34	100.0
2.	Ampicillin/Sulbactam(10/10µg)	A/S	34	100.0
3.	Ticarcillin(75µg)	TC	34	100.0
4.	Piperacillin/Tazobactam(100/10µg)	PIT	28	82.4
5.	Cefotaxime(30µg)	CTX	34	100.0
6.	Ceftriaxone(30µg)	CTR	34	100.0
7.	Ceftazidime(30µg)	CAZ	34	100.0
8.	Cefepime(30µg)	CPM	34	100.0
9.	Cefepime/Tazobactam(30/10 µg)	CPT	33	97.1
10.	Cefoperazone/Sulbactam	CFS	30	88.2
11.	Aztreonem	AT	34	100
12.	Amikacin(30µg)	AK	27	79.4
13.	Gentamycin(10µg)	GEN	29	85.3
14.	Tobramycin(10µg)	TOB	33	97.1
15.	Netilmicin	NET	32	94.1
16.	Ciprofloxacin(5µg)	CIP	30	88.2
17.	Ofloxacin(1 µg)	OF	29	85.3
18.	Levofloxacin(5µg)	LE	28	82.4
19.	Tetracycline(30µg)	TE	29	85.3
20.	Cotrimoxazole(1.25/23.75µg)	COT	32	94.1
21.	Imipenem(10µg)	IMP	27	79.4
22.	Meropenem(10µg)	MRP	27	79.4
23.	Polymyxin B	PB	0	0.0
24.	Tigecycline	TGC	0	0.0

Table 3: Different methods of detection of Carbapenem susceptibility

	Screening test	MHT	CDDT	Genotypically
Carbapenem resistance	27	11	18	23

Table 4: Diagnostic test evaluation; Sensitivity and specificity of MHT and CDDT

	MHT	CDDT
Sensitivity	47.83%	78.26%
Specificity	100%	100%
Positive predictive value	100%	100%
Negative predictive value	47.83%	68.75%

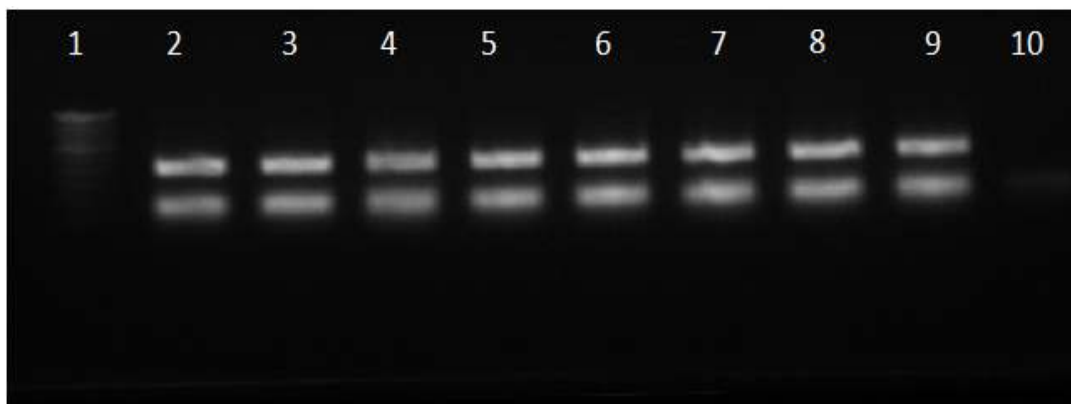


Fig. 1: The photographs show positive confirmation of bla_{NDM-1} gene from lane 2 to 9 were found positive samples, lane 10 shows negative result and lane 1 associated with 100bp ladder

DISCUSSION

Non fermenting Gram-negative bacteria *Acinetobacter baumannii*, widely associated with long term care facilities. In this study 200, test cases were studied from six different hospitals of Kanpur and 315 samples were collected and microbiologically processed.

In this study total of 200 patients were observed for nosocomial infections, admitted to ICU of different tertiary care hospitals and total 34 *Acinetobacter baumannii* was isolated. Species *A. baumannii* was confirmed by presence of bla_{OXA 51} gene. On the basis of microbiological and clinical correlation among 34 isolated *A. baumannii*, only 20 were contributed to the infection. The rate of *Acinetobacter* infection was 9.18 per 1000 ICU days.

Omer et al reported approx similar result, 9.5% of *Acinetobacter* infection in their study (Omer et al., 2015). There were some another studies, in Rajasthan 83.2% of *A. baumannii* was reported as the most common nosocomial pathogen especially from ICUs (Sharma et al., 2013). Similarly, in Pune, *Acinetobacter* was the most common isolates from ICU (Patwardhan et al., 2008). The differences among the studies can be attributed to differences in study populations and locality, availability of resources, overcrowding or shortage of nurseries, antibiotic use and the difference in surveillance methods for detection of nosocomial infections.

Here only polymyxins and tigecycline was found to be 100% sensitive other studies showed also

that these are the most effective drugs (Urban et al., 2003 and Jaggi et al., 2012). This study brings up an important aspect of increasing resistance in *A. baumannii* towards carbapenems and harbor bla_{NDM-1}.

The increasing reports on the NDM-1 producing bacteria are a major concern worldwide. Although, NDM-1 seems to be endemic in the Indian subcontinent, other studies have suggested that the Balkan countries may act as another reservoir of the NDM-1 producers (Kumarasamy et al., 2010 and Lascols et al., 2009)

Here, we are reporting the presence of NDM-1 producing *A. baumannii* isolates in various clinical samples from a tertiary care hospital. The overall prevalence of the bla_{NDM-1} possessing *A. baumannii* isolates in Kanpur was found to be 5.5% (11/200).

Screening for the drug susceptibility reported 27 strains were carbapenem resistant. Phenotypically only 18 strains were found to produce carbapenemases by CDDT, while only 11 strains were MHT positive.

MHT has been non specific for the detection of metallo-beta-lactamases (Thomson et al., 2010) in the present study MHT detected 6 of the 11 NDM-1 positive isolates. The CDDT detected 9 of the 11 isolates that were positive for the bla_{NDM-1} gene. Therefore only molecular tools were capable of detecting NDM in *A. baumannii*, and it is likely that

carbapenem resistance is the result of decreased production of outer membrane porins, together with low-level expression of NDM (Naas et al., 2009 and Poirel et al., 2006).

In this study, samples were collected from ICU. Among the 11 bla_{NDM-1} positive *A. baumannii* isolates, a majority of the infected patients were died (5/11).

Bonnin et al. recently suggested that *A. baumannii* not only might accept resistance genes but also could act as a gene donor, spreading resistance genes to other bacteria, including Enterobacteriaceae (Bonnin et al., 2014). This possibility emphasizes our concerns about dissemination of these genes in our country as in many other countries where NDM-1-producing *A. baumannii* have been isolated, making these findings a global public health matter, as suggested by Johnson and Woodford (Bonnin et al., 2014).

CONCLUSION

Carbapenem Resistant *A. baumannii* was prevalent in our Kanpur region. Carbapenemase genes bla_{NDM} in clinical isolates of *A. baumannii* is a worrying trend and it underlines the need for a stringent infection control practices along with antimicrobial stewardship to curb these *Acinetobacter* species.

ACKNOWLEDGEMENTS

The authors are thankful to the Principal and HOD (Microbiology) of the Rama Medical College Hospital and Research Centre, Kanpur (India) for the providing facilities and permission to carry out the work.

REFERENCES

Yong D., Toleman M. A., Giske C. G., Cho H. S., Sundman K., Lee K., et al., 2009. Characterization of a new metallo-beta-lactamase gene, bla_{NDM1}, and a novel erythromycin esterase gene which was carried on a unique genetic structure in *Klebsiella pneumoniae*, sequence type 14 from India. *Antimicrob Agents Chemother*, **53**(12):5046-5054.

Kumarasamy K. K., Toleman M. A., Walsh T. R., Bagaria J., Butt F., Balakrishnan R., et al., 2010. Emergence of a new antibiotic resistance mechanism in India, Pakistan, and the UK; A molecular, biological, and epidemiological study. *Lancet Infect Dis*, **10**(9):597-602.

Salcido R. S., 2010. Super bugs: survival of the fittest. *Adv Skin Wound Care*, **23**(10):439.

Pal N., Sujatha R., Kumar A., 2017. Phenotypic and Genotypic Identification of *A. baumannii* with special reference to Bla_{oxa-51}like gene and its antimicrobial susceptibility pattern from intensive care units in Kanpur. *International Journal of Contemporary Medical Research*, **5**:1154-1158.

CLSI, 2016. Performance standard for Antimicrobial susceptibility testing; twenty third informational supplement. M100-S26, **33**(1).

Omer M. I., Gumaa S. A., Hassan A. A., Idris K. H., Ali O. A., Osman M. O., Saleh M. S., Mohamed N. A., Khaled M. M., 2015. Prevalence and resistance profile of *Acinetobacter baumannii* clinical isolates from a private hospital in khartoum, sudan. *American Journal of Microbiological Research*, **3**(2): 76-79.

Sharma D. K. , Tiwari Y. K., Vyas N. and Maheshwari R. K., 2013. An investigation of the incidence of nosocomial infections among the patients admitted in the intensive care unit of a tertiary care hospital in Rajasthan, India. *Int. J. Curr. Microbiol. App. Sci*, **2**(10):428-435.

Patwardhan R. B., Dhakephalkar P. K., Niphadkar K. B., Chopade B.A., 2008. A study on nosocomial pathogens in ICU with special reference to multi-resistant *Acinetobacter baumannii* harbouring multiple plasmids. *Indian J. Med. Res.*, **128**(2):178-87.

Urban C., Segal-Maurer S., Rahal J. J. 2003. Considerations in control and treatment of nosocomial infections due to multidrug

- resistant *Acinetobacter baumannii*. Clin. Infect. Dis., **36**:1268-1274.
- Jaggi N., Sissodia P., Sharma L., 2012. *Acinetobacter baumannii* isolates in a tertiary care hospital: Antimicrobial resistance and clinical significance. J. Microbiol. Infect. Dis., **2(2)**:57-63.
- Lascols C., Hackel M., Marshall S. H., Hujer A. M., Bouchillon S., Badal R., et al., 2009. The increasing prevalence and the dissemination of NDM-1 metallo- β -lactamase in India: data from the SMART study. J. Antimicrob. Chemother., **66(9)**:1992-1997.
- Thomson K. S., 2010. Extended-spectrum β -lactamase, Amp C, and 16. carbapenemase issues. J Clin Microbiol, **48**: 1019-25.
- Naas T., Nordmann P., 2009. *Acinetobacter* and β -lactams, p. 407-420. In Courvalin P., Le Clercq R., Rice L. B. (ed), Antibioqram, 3rd ed. ASM Press, Washington, DC.
- Poirel L., Nordmann P., 2006. Carbapenem resistance in *Acinetobacter baumannii*: mechanisms and epidemiology. Clin. Microbiol. Infect, **12**: 826-836.
- Bonnin R., Poirel L., Nordmann P., 2014. New Delhi metallo- β -lactamase-producing *Acinetobacter baumannii*: a novel paradigm for spreading antibiotic resistance genes. Future Microbiol., **9**:33-41.