## BIOFOULING ORGANISMS ATTACHED ON MANGROVE PLANTS IN AYIRAMTHENGU REGION, KERALA, INDIA

# BINSY M. KESAVAN<sup>a1</sup> AND S. AMINA<sup>b</sup>

<sup>ab</sup>Research Department of Zoology, S.D. College, Alappuzha, University of Kerala, India
<sup>b</sup>M.S.M College, Kayamkulam, University of Kerala, India

## ABSTRACT

Epibionts that colonize on the mangrove areial roots and stem was analysed during February 2016 to January 2017 covering all the seasons. Micro and macro foulers were attached to the mangrove plants. *Balanus sp.* was dominant macrofouler while *Conscinodiscus sp, Pleurosigma sp, Chlorella sp*, were the abundant microfoulers. Maximum settlement was on the premonsoon period and poor during monsoon season. Hydrological parameters showed a seasonal variation throughout the study period. Temperature ranges from 27.  $5^{\circ}$ C to  $30^{\circ}$ C, dissolved oxygen (4. 6 to 5. 8mg/l), salinity(19. 8 to 32. 7ppt). The epibionts exhibited distinct seasonal trends in abundance with fluctuating temperature and salinity. Species richness, abundance and diversity was higher during premonsoon and lower on monsoon period.

KEYWORDS: Epibionts, Mangrove, Microfouler, Macrofouler, Species Richness

Biofouling is the undesirable accumulation of sessile micro and macro organism with time (Wahl, 1987; Railkin 2004; Durr and Thomason 2010). The species composition of fouling community change considerably over space for biofouling is inferred as the process of accumulation of micro-organism, diatoms algae, plants and animals on wetted surface. Fouling occurs world wide in various industries, shiphull, underwater equipments beach well structures (Haderlie, 1981) and also in the roots of mangrove plants (Karuppayian and Raja, 2007; Rani et al 2010). Fouling is not a simple process its sequence and specific composition depends upon location, year, seasons and various physicochemical factors (O' Neil & Wilcox 1971).

Extensive work has been done all over the world on understanding biofouling on mangroves and their deleterious effect (Cancinni et al, 2008). Biofouler form thick hard encrustation on roots, branches, leaves and stem of the mangrove. Epibionts are found on world wide on mangrove roots (Rutzler 1969, Alvare Z I, 1989; Ellison 1992). Marine tidal forest, the mangroves cover about 137,800 sqkm of the world's tropical and sub-tropical region. In Kerala total extent of mangrove is in 2502 ha (Mini et al 2012). Researches has been carried out in Ayiramthengu mangrove to study the various aspects; but this is the first report of biofouling in the mangrove roots and stem from Kerala. Preliminary studies on epibiotic protist in the mangrove ecosystem of Ayiramthengu showed that 15 species were identified from the prop roots of R. apiculata (Chitra and Sunil, Kumar, 2015). Oyster infestation on mangroves are reported from Vietnam (Hoang and Nhuong, 2004), Africa (Gillikin and Verheyden, 2002), Hong Kong (Chiu 2000) and India

(Rani et al, 2010, Rajani Panchang, 2014). The present study was to analyze the biofouling (epibionts) on the roots and stem of mangroves plants in Ayiramthengu region during February 2016 to January 2017.

## MATERIALS AND METHODS

#### **Study Area**

Ayiramthengu Mangrove (9<sup>0</sup> 6' 8'' N Latitude to 76<sup>0</sup> 28' 29'' E Longitude) is situated on the Kollam district, Kerala, India. The mangrove ecosystem is a part of Kayamkulam estuary which is a narrow stretch of tropical backwater on the west coast of Peninsular India. Mangrove is dominated by *Avicennia marina* and *Rhizopora apiculata*. This mangrove was declared as an Environmental Hot Spot after it was ravaged by 2004 Indian Ocean Tsunami and threatened with extinction due to human interference.

#### **Collection of Samples**

Fouling organism were scraped and from the mangrove roots and stem of *A. marina* and *R.apculata*, kept in plastic bottles. The samples were brought to the laboratory for further analysis. The microfoulers were identified by using compound microscope its number were noted and recorded. The macrofoulers were identified from the site itself and its number recorded. To assess the diversity of the epibionts indices like species richness, species abundance, Simpson dominance, species diversityindex(Shannon Weiner) were used. Water samples were collected for the analysis of hydrological parameters like temperature, dissolved oxygen and salinity by using standard methods (APHA, 2005).

### **RESULTS AND DISCUSSION**

Biofouling organisms on the mangrove roots of Ayiramthengu mangrove during the premonsoon, monsoon and post monsoon were collected and identified. The abundance of the fouling organisms were subjected to geographical and seasonal variation. For this water quality parameters like temperature. dissolved oxygen and salinity were calculated and the mean temperature ranges from  $27.5^{\circ}$ C to  $30^{\circ}$ C. Dissolved oxygen and salinity depicts the values 4.6 mg/l, 5.8 mg/l, 5.2 mg/l and 32.7 ppt, 19.2 ppt, 29 ppt during the premonsoon, monsoon and post monsoon respectively (Table 1).

The surface water temperature is influenced by

the density of solar radiation, evaporation freshwater

influx, cooling and mixup with ebb and flow from

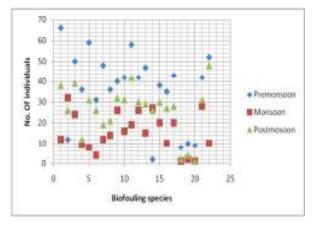
adjoining neretic waters. (Santhakumaran and Sawant,

1998). Balanus sp. is a predominant fouler of mangrove

Sl.No	Parameters	Premonsoon	Monsoon	Postmonsoon
1.	Temperature ( <sup>0</sup> C)	30	27.5	28.2
2.	Dissolved Oxygen (mg/l)	4.6	5.8	5.2
3.	Salinity (ppt)	32.7	19.2	29

Table 1: The Physico-chemical parameters of Ayiramthengu Mangrove during February 2016 to January 2017

The epibionts belongs to Phylum Ochrophyta includes the Class Bacillariophyceae comprises 15 Chlorophyta genus. Phylum with one class Trebouxiophyceae constitute the genus Chlorella sp. and class Hormogoneae of Cyanobacteria with the Anabaena sp. The macrofouling, species is from the Phylum constitute class Arthropoda Malacostraca and Maxillopoda with the genus Gammarus sp. and Balanus sp. respectively. Altogether the epibionts of the mangrove roots and stem belongs 5 phylum and 6 classes. The abundant microfouling species are Conscinodiscus sp. Pleurosigma sp. and Chlorella sp. during premonsoon period and macrofouler was Balanus sp.(Table 2,Graph 1) The Species richness (22) dominance (0.944, 0.938, 0.941) species abundance (806, 336, 573) of the epibionts during premonsoon, monsoon and postmonsoon respectively. Shannon Weiner Diversity depicts the values 2.995, 2.884 and 2.932 on premonsoon, monsoon and postmonsoon period Table (3).



roots and stem. Their settlement was high during late post monsoon and early premonsoon period due to optimal temperature and salinity for the growth and breeding. Barnacle showed an increasing trend gradually during January and March (Karuppayian and Raja, 2007). Barnacles were present through out the year and were absent in high tide level on mangroves. During low tide level on premonsoon and later during post monsoon there were mangrove settlement. The effects of barnacles occurred erratically on stem seedlings and on leaves had little effect on the growth of mangrove plants. The calcareous base for attachment of this species damage the plants (Satumanatpan and Michael J Keough, 1999). The settlement and distribution of barnacle on mangrove is affected by factors like salinity, depth, canopy, hydrological regime and biotic factors like aminoacid composition and one dimensional structure of barnacle adhesive protein (Xiang et al, 2006). Here the surface water temp, salinity influences the Balanus sp. distribution of on mangrove roots and stem. In the mangrove species Sonneratia apetala the leaves possess possible anti barnacle defence strategies suggesting that it was less frequent and less abundantly fouled by barnacles (Danquing et al, 2016).

Graph 1: showing the distribution of biofouling species collected from mangroves

No	Species	Premonsoon	Monsoon	Postmosoon
1.	Conscinodiscus	66	12	38
2.	Melosira	12	32	26
3.	Nitzschia	50	24	39
4.	Cymbella	36	9	12
5.	Pleurosigma	59	8	31
6.	Licmophora	31	4	26
7.	Ceratium	48	12	19
8.	Cocconeis	36	14	21
9.	Navicula	40	26	32
10.	Diatoma	42	16	31
11.	Chorella	58	19	42
12.	Rhizosolenia	42	26	30
13.	Leptocylidricus	47	15	29
14.	Anabena	2	27	26
15.	Thalassionema	38	20	30
16.	Biddulphia	35	10	27
17.	Chaetoceros	43	20	28
18.	Golbigerina	8	1	2
19.	Elphidium	10	2	4
20.	Textularia	9	1	1
21.	Gammarus	42	28	31
22.	Balanus	52	10	48

Table 2: Biofouling organisms collected from mangrove roots and stem from Ayiramthengu region duringFebruary 2016 to January 2017.

Table 3: Diversity indices of biofoulers collected from the roots and stem of mangrove plants	<b>Table 3: Diversit</b>	v indices of biofoulers col	lected from the roots and	stem of mangrove plants
---	--------------------------	-----------------------------	---------------------------	-------------------------

Diversity indices	Premonsoon	Monsoon	Postmonsoon
Species richness	22	22	22
Species abundance	806	336	573
Dominance	0.944	0.938	0.941
Species Diversity	2.995	2.844	2.923

The increasing dissolved oxygen in the mangrove is caused due to oxygen exchange at the root system and the flow of dissolved oxygen enriched inland water(Rahman et al,2007). Biofouling on roots cause adverse effect to the mangroves. Postmonsoon favours the settlement of biofoulers. The influx of freshwater and rainwater into the mangrove affect the salinity variation (Kesavan et al, 2013). Salinity was mainly determined by local hydrology where input of salt water from periodic tides and fresh water from rivers, rainfall and runoff (Jadhav, 2011). Low salinity during monsoon favours the settlement of *Melosira sp* and *Anabaena sp*. The phytoplankton microfoulers is high during premonsoon period. The seasonal distribution depends greatly on the degree of variation in salinity and temperature. The fresh water phyto plankton *Melosira sp.* and *Anabaena sp.* showed its abundance during low temperature and salinity (Mitbavkar and Anil, 2008)

On conclusion although 22 species of epibionts including micro and macrofoulers were noticed during the present study. The colonization of epibionts on the roots, stems and leaves may adversely affect their growth. The hydrological parameters, wave exposure to roots and stem influence the biofouling distribution and diversity on the Ayiramthengu mangrove during February 2016 – January 2017. Biofouling in the roots and stem of mangrove plants showed that increase in species richness and abundance with increase in temperature and salinity. Shannon Weiner

diversity index showed higher value during premonsoon indicates that diversity is maximum. *Balanus sp.* was the predominant fouler in the mangrove, necessary steps are taken to protect the mangrove trees from the biofouling organisms, especially from the *Balanus* species.

### ACKNOWLEDGEMENT

The authors are thankful to the authorities of 27<sup>th</sup> Swadesi Science Congress for giving the opportunity.

## REFERENCES

- Aaron M. Ellison and Elizabeth J. Farnsworth, 1992. The ecology of Belizean mangrove-root fouling communities: patterns of epibiont distribution and abundance, and effects on root growth. The Ecology of Mangrove and Related Ecosystems, Springer, pp 87-98.
- Alvarez I.A., 1989. Establecimiento, desarrollo y mantenimiento de una comunidad epibentonica tropical. Ph.D. Thesis, Universidad Central de Venezuela, Facultad de Ciencias. 150 pp.
- APHA, 2005. Standard Methods for the Examination of Water and Wastewater. 21st Edition, American Public Health Association/American Water Works.
- Cancinni S., Burrows D., Fratini S., Smith T.J., Offenberg J. and Guebas F.D., 2008. Faunal impact on vegetation structure and ecosystem function in mangrove forests: A review. Aqua. Bot., 80:186-200.
- Canning-Clode J. and Martin Wahl, 2010. Patterns of Fouling on a Global Scale. Blackwell Publishing Ltd.
- Chitra and Kumar S., 2015. Preliminary studies on epibiotic protist in mangrove ecosystem of ayiramthengu, Kerala Coast.
- Chiu H.M.C., 2000. The Distribution and shell allometry of the Rock Oyster, in The Marine Flora and fauna of Hong Kong and Southern China, edited by B. Morton, Proceedings of the 10th International Marine Biological Workshop, (Hong Kong University Press), pp. 331-340.
- Danqing Feng, Wei Wang, Xiang Wang, Yan Qin and Cai-huan Ke, 2016. Low Barnacle fouling in the leaves of mangrove plant *Sonneratia apetala* and the possible anti-barnacle defense strategies. Marine Ecology Progress Series, **544**:169-182.

- Durr S. and Thomason J.C., 2010. Biofouling. Wiley-Blackwell, Oxford, UK.456pp.
- Gillikin D. and Verheyden A., 2002. Field Guide to Kenyen Mangroves.
- Haderlie E.C., 1981. Technology for control of marine biofouling- a review, Naval Institute Press, Annapolis, 163.
- Hoang N.K. and Nhuong D.V., 2004. Some results of research on benthos attached to mangroves and their adverse impacts, in Mangrove Ecosystems in the Red River Coastal Zone edited by P.N. Hong, (Agricultural Publishing House, Hanoi, Vietnam).
- Jadhav R., 2011. News-article- Bharnaare pot hi ritech! Loteteel vikaasachi vishaari vilkhabhaag 4. Pudhari, 16<sup>th</sup> May 2011.
- Karuppaiyan M. and Raja K., 2007. Effect of Tides on Settlement of Oysters and Barnacles in Pichavaram Mangrove of the Southeast Coast of India. Res. J. Environ. Sci., 1:324- 330.
- Kesavan K.C., Palpandi T., Amalraj S., Vairamani S., Raveendran and Shanmugam A., 2007.
  Population density and distribution pattern of malacofauna in the mangroves along Coringi river (Godavari mangroves, Andhra Pradesh) National Conference Emerging Scenario in Zoology 27<sup>th</sup> Sept, pp.70-76.
- Martin Wahl, 1987. Marine epibiosis. I. Fouling and antifouling: some basic aspects, Marine ecology progress series, pp.175-189.
- Mohandas M., Lekshmi S. and Radhakrishnan T., 2012. Kerala Mangroves – Pastures of Estuaries : Their present status and challenges. International Journal of Science and Research (IJSR) ISSN (online): 2319-7064, pp 2804-2809.
- O' Neil T.B. and Wilcox C.L., 1971. The formation of primary film on materials submerged in the sea at Port Hueneme, California, Pacific Science, **25**(1):1-12.
- Rahaman S.M.B., Sarder L.M. S., Rahaman A.K., Ghosh S.K., Biswas S.M.S., Siraj K.A., Huq A.F., Hasanuzzamann M. and Islam S.S., 2013. Nutrient dynamics in the Sundarbans mangrove estuarine system of Bangladesh under different weather and tidal cycles, Ecol. Proc., 2:29.

- Railkin A.L., 2004. Marine Biofouling: Colonization Process and Defenses. CRC Press, Boca Raton, Florida, 303 pp.
- Rajani P., 2014. Biofouling by Saccostrea cucullata a major threat to mangroves of Vasishthi Estuary, Maharashtra , Indian Journal of Geo-Marine Science, 44(8):1155-1161.
- Rani S.S., Bragadeeswaran K.P. and Priyadarshini S., 2010. Infestation of Barnacle (*Balanus amphitrite*) in the Mangrove Environment of Vellar Estuary, Tamilnadu, World J. Fish Mar. Sci., 2(4):307-310.
- Rutzler K., 1969. The mangrove community, aspects of its structure, faunistics and ecology. In Lagunas

Costeras, un Simposio. UNAM-UNESCO, Mexico, pp.515–535.

- Santhakumaran L.N. and Sawant S.G., 1998. Marine Wood – infesting organisms in the destruction of living mangrove vegetation along Goa coast. Indian Forest Bulletin, ICFRE, Dehradun, 67 pp.
- Satumanatpan S. and Michael J. Keough, 1999. Effect of Barnacles on the survival and growth of temperate mangrove seedlings. Mar. Ecol. Prog. Ser., **181**:189-199.
- Xiang P., Yang Z. and Lin P., 2006. Barnacle damage and its control in young mangrove plantation: A research review. NCBI Pub Med. Gov., **17**(8): 1526-29.